

META ANALYSIS

High-Risk Human Papillomaviruses in Oral Squamous Cell Carcinoma (OSCC): A Meta-Analysis

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ABSTRACT

Background: The prevalence of high-risk Human Papillomaviruses (HPV) in cases of oral cavity squamous cell carcinoma (OSCC) varies widely. Therefore, the aim of this study was to determine the pooled prevalence of all high-risk HPV by meta-analysis with specific emphasis on HPV type 16/18.

Methods: The studies were retrieved from PubMed and MEDLINE to conduct a comprehensive literature review on HPV detection in OSCC. Search terms included, High-risk HPV, oral cancer, polymerase chain reaction (PCR), in situ hybridization (ISH). We reviewed 47 research studies systematically to report the prevalence of high-risk HPV infection in oral cancer. Included studies published from 1988 to 2018. The meta-analysis was carried out by using MedCalc software version 19.0.3.

Results: A meta-analysis was executed to calculate the pooled prevalence of High-risk HPV types, which revealed overall decreasing order frequency of high-risk HPV and high-risk type displaying the highest number of type 16/18 HPV in the reported cases. As 30.71% [24.59 to 37.19 % confidence interval (CI) at 95%] and 28.88 % [22.62 to 35.57% confidence interval (CI) at 95%] followed by other high-risk HPV 3.59% [2.22 to 5.46%] respectively.

Conclusions: According to present meta-analysis, we conclude that 16/18 HPV displaying maximum infection rate as compared to other high-risk HPV type in OSCC cases.

Keywords: High-Risk HPV Detection; PCR; Oral Cancer; Meta-analysis.

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INTRODUCTION

The term "Oral cancer" includes neoplasms affecting pharyngeal regions, salivary glands and oral cavity. However, the term "Oral cancer" has a propensity to be used with Oral squamous cell carcinoma (OSCC), which implies the most common of all oral neoplasms. About 90% of all oral neoplasms are Oral squamous cell carcinoma (OSCC)¹. This is the sixth most common cancer in the world^{2,3}. It accounts for 2-4 % of all cancers globally, with major incidence in western countries⁴⁻⁶.

With the available therapeutic approaches, there is no improvement in the morbidity and mortality rates of OSCC in the past 30 years⁷. Alcohol, tobacco and smoking are the major independent risk factors

in the escalating prevalence of oral cancer; however, their action seems to be combined. The combinations of tobacco and alcohol consumption possess a synergistic effect on the development of oral cancers⁸. However, tobacco alone is one of the most important escapable risk factor that is responsible for one-fourth of cancer deaths worldwide^{2,9}. Other risk factors that contribute to the progression of oral cancers include betel quid chewing, areca nut, narcotics and cannabis^{10,11}. A part from these risk factors Human Papillomavirus (HPV) has also been implicated to positively associate with oral pre-cancer and cancerous lesions^{12,13}. Syrjanen et al. first reported HPV involvement in oral and oropharyngeal carcinogenesis in 1983^{1,4}. A sound upsurge of HPV incidence in oral cancer cases has been observed over the last few

decades. It has been positively associated with OSCC prognosis and young age onset in several developed countries¹⁴.

The human papillomavirus (HPV) belongs to the family of DNA Papovaviridae. It consists of double-stranded DNA (approximately 8,000 base pairs long) which is enclosed in a protein capsid¹³. The HPV genome consists of a long non-coding region (LCR), six early and two late open reading frames (ORFs)¹⁵. Viral replication and transcription of early genes are controlled by the long non-coding region¹³. The two key viral proteins, E6 and E7 play an important role in the transformation and immortalization of the cell by inactivating p53 and Rb genes¹⁶. More than 200 HPV genotypes have been sequenced out of which 85 are well-characterized¹⁶. High-risk oncogenic types, HPV 16 and HPV 18 are more closely associated with oropharyngeal cancers^{17,18}.

In recent years, many researchers have focused on the relationship between HPV and OSCC. However, significant heterogeneity exists in the literature regarding OSCC and HPV frequency. The difference among geographical locations as well as HPV detection method could be the key contributors of the variation. Therefore, it is crucial to implement a meta-analysis to evaluate comprehensively the relationship between high risk HPV and OSCC. The HPV prevalence rate in oral cancer was found to be varied from 0% to 100%^{6, 13, 32}. The main objective of this study was to determine the pooled prevalence of all high-risk HPV including type 16/18 HPV in OSCC patients by meta-analysis.

METHODS

Source of Data

The PubMed and MEDLINE, databases were used to conduct a comprehensive literature review on HPV detection in OSCC. English language studies were considered for literature search, which were published between 1988 and 2018. Search terms included, High-risk HPV, oral cancer, polymerase

chain reaction (PCR), in situ hybridization (ISH). Where, 5 studies of ISH and 42 studies of PCR are included.

Results of Search Strategies

Out of 205 publications related to search strategy, those articles, which were in other language (n=18), head and neck cancer (n=36), citation do not relate with this study (n=52), unreliable detection methods (n=15), non-uniformity of data (n=35) were excluded. Based on the initial review, 47 studies that reported HPV prevalence in OSCC were included in this meta-analysis.

Statistical Analysis

The prevalence of high-risk HPV (16, 18, 31, 33, 35, 52 and 56) and 95% confidence intervals (CI) were calculated for each study. Forest plots were constructed to carry out meta-analysis by using MedCalc software version 19.0.3. Overall prevalence was defined as the number of OSCC samples that tested positive for high risk HPV divided by the total number of OSCC samples. We explored the heterogeneity between study-specific results using the chi-square-based Cochran's Q test. Estimates were obtained by random-effects models for overall high-risk HPV and HPV 16/18 type and the fixed-effects model for other high-risk HPV (31, 33, 35, 52 and 56).

RESULTS

Meta-analysis for pooled prevalence based on all high-risk HPV type

The 47 studies evaluated a total of 3775 OSCC cases. The number of cases varied from 13 to 254 (Table 1). The detection rate of high-risk HPV was different based on study and the method of detection. Included studies reported the prevalence rate of high-risk HPV types. On the basis of random effect model the prevalence of high-risk HPV types in OSCC was 30.71% [24.59 to 37.19% confidence interval (CI) at 95%]; the significant heterogeneity ($p < 0.0001$) was found in the studies [I² = 94.46% with 93.35 to 95.38% confidence interval (CI) at 95%].

Table 1: Prevalence of High-Risk HPV in Oral Cancer based on 47 studies.

Study	Sample size	Proportion (%)	95% CI	Weight (%)
Syrjanen et al.(1988) ⁴⁰	51	11.765	4.442 to 23.868	2.12
Chang et al.(1990) ⁴¹	40	2.500	0.0633 to 13.159	2.07
Chang et al.(1990) ⁴¹	40	27.500	14.601 to 43.888	2.07
Holladay and Gerald (1993) ⁴²	39	17.949	7.535 to 33.535	2.07
Brandwein et al.(1994) ⁴³	64	25.000	15.016 to 37.399	2.15
Van Rensburg et al.(1995) ⁴⁵	66	1.515	0.0384 to 8.155	2.16

Shindoh et al.(1995) ⁴⁴	77	31.169	21.095 to 42.743	2.18
Balaram et al.(1995) ³¹	91	41.758	31.501 to 52.567	2.20
Chiba et al(1996) ⁷⁴	38	21.053	9.554 to 37.319	2.06
Cruz et al.(1996) ⁴⁶	35	54.286	36.646 to 71.173	2.04
Wen et al.(1997) ⁴⁷	45	31.111	18.166 to 46.649	2.09
Premoli -De-Percoco et al.(1998) ⁴⁸	50	70.000	55.392 to 82.138	2.11
Schwartz et al.(1998) ⁴⁹	193	21.244	15.697 to 27.696	2.25
Pillai et al.(1999) ⁵⁰	61	27.869	17.147 to 40.829	2.15
Cao et al (2000) ⁵²	40	72.500	56.112 to 85.399	2.07
Patima et al. (2000) ⁵³	73	73.973	62.376 to 83.546	2.17
Gillison et al.(2000) ⁶⁹	84	11.905	5.859 to 20.805	2.19
Bouda et al.(2000) ⁵¹	19	94.737	73.972 to 99.867	1.87
Premoli -De-Perco et al.(2001) ⁵⁴	50	60.000	45.179 to 73.592	2.11
Shima et al.(2000) ⁷⁷	44	20.455	9.804 to 35.305	2.09
Schwartz et al.(2001) ⁵⁵	254	15.748	11.495 to 20.821	2.27
Nagpal et al. (2002) ⁵⁶	110	33.636	24.908 to 43.271	2.22
Kumar et al. (2003) ⁵⁸	42	30.952	17.622 to 47.086	2.08
Sugiyama et al. (2003) ⁶⁰	86	34.884	24.919 to 45.923	2.19
Chang et al. (2003) ⁵⁷	103	49.515	39.514 to 59.544	2.21
Ritchie et al. (2003) ⁵⁹	141	14.894	9.462 to 21.861	2.24
Zhang et al. (2004) ⁶⁴	73	73.973	62.376 to 83.546	2.17
Correnti et al. (2004) ⁶¹	16	50.000	24.651 to 75.349	1.81
Smith et al. (2004) ⁶³	106	9.434	4.617 to 16.666	2.21
Dahlgren et al. (2004) ⁶²	110	10.909	5.765 to 18.281	2.22
Ibieta et al.(2005) ³⁵	21	66.667	43.032 to 85.412	1.90
Boy et al.(2006) ⁶⁵	59	11.864	4.906 to 22.929	2.14
El -Mofty et al. (2006) ⁶⁶	94	34.043	24.581 to 44.541	2.20
Badaracco et al.(2006) ⁷⁰	60	13.333	5.936 to 24.592	2.14
Nemes et al.(2006) ⁶⁷	79	41.772	30.767 to 53.413	2.18
Sugiyama et al. (2007) ⁶⁸	27	37.037	19.401 to 57.632	1.98
Smith et al. (2008) ⁷¹	108	24.074	16.368 to 33.251	2.21

Kong et al.(2009) ⁷²	13	7.692	0.195 to 36.030	1.73
Chaudhary et al.(2010) ³⁷	208	34.615	28.171 to 41.505	2.26
Attner et al.(2010) ⁷³	87	78.161	68.015 to 86.310	2.19
Gonzalez-Ramirez I et al.(2013) ³⁴	80	5.000	1.379 to 12.310	2.18
Gan et al.(2014) ³³	200	19.500	14.249 to 25.679	2.26
Koukertsu A et al.(2016) ³⁹	24	54.167	32.821 to 74.447	1.94
Chen et al.(2016) ³⁶	178	3.371	1.247 to 7.192	2.25
Palve et al.(2018) ³⁸	106	33.019	24.190 to 42.824	2.21
Rubab Z et al. (2018) ⁷⁵	100	32.000	23.022 to 42.077	2.21
De Abreu PM (2018) ⁷⁶	90	3.333	0.693 to 9.434	2.20
Total (Random Effects)	3775	30.713	24.593 to 37.195	100.00

Test for heterogeneity

Q	830.0819
DF	46
Significance level	P < 0.0001
I2 (inconsistency)	94.46%
95% CI for I2	93.35 to 95.38

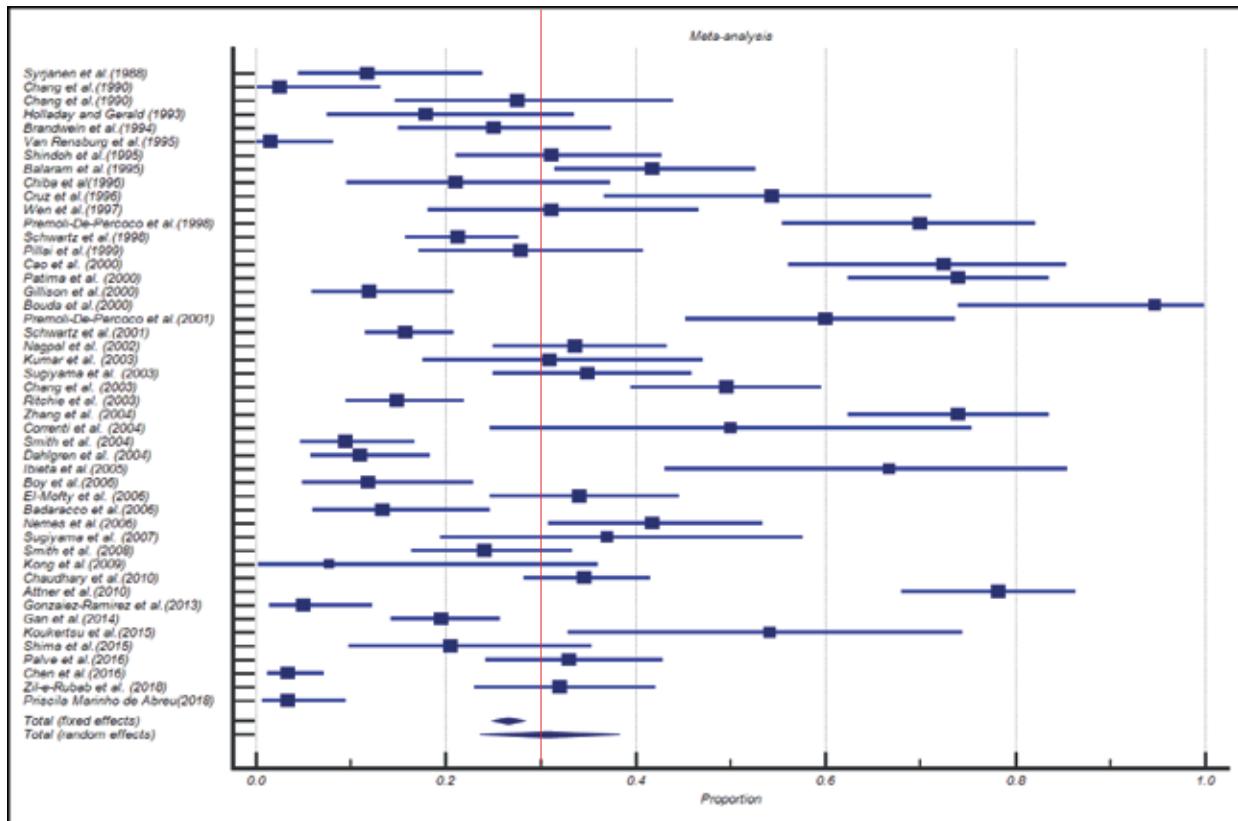


Figure 1: Forest Plot for Prevalence of High-Risk HPV in Oral Cancer.

Meta-analysis for overall prevalence based on HPV 16/18 type

Of the 47 research studies, 38 studies included to calculate pooled prevalence of HPV- 16/18 in OSCC. The significant heterogeneity ($p < 0.0001$) was found among the studies [$I^2 = 93.96\%$ with 92.57 to 95.10% confidence interval (CI) at 95%]. Out of

3158 patients, 850 patients were positive for HPV-16/18. Proportion of positive cases for HPV 16/18 was also analyzed in the present meta-analysis. Based on random effect model the mean proportion of positive cases was 28.88% [22.62 to 35.57% confidence interval (CI) at 95%].

Table 2: Prevalence of HPV 16/18 in Oral Cancer based on 38 studies.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)
Syrjanen et al.(1988) ⁴⁰	51	11.765	4.442 to 23.868	2.59
Chang et al.(1990) ⁴¹	40	2.500	0.0633 to 13.159	2.53
Chang et al.(1990) ⁴¹	40	27.500	14.601 to 43.888	2.53
Holladay and Gerald (1993) ⁴²	39	17.949	7.535 to 33.535	2.52
Brandwein et al.(1994) ⁴³	64	25.000	15.016 to 37.399	2.64
Van Rensburg et al.(1995) ⁴⁵	66	1.515	0.0384 to 8.155	2.65
Shindoh et al.(1995) ⁴⁴	77	31.169	21.095 to 42.743	2.68
Balaram et al.(1995) ³¹	91	41.758	31.501 to 52.567	2.70
Cruz et al.(1996) ⁴⁶	35	54.286	36.646 to 71.173	2.49
Wen et al.(1997) ⁴⁷	45	31.111	18.166 to 46.649	2.56
Premoli -De-Percoco et al.(1998) ⁴⁸	50	70.000	55.392 to 82.138	2.59
Schwartz et al.(1998) ⁴⁹	193	21.244	15.697 to 27.696	2.78
Pillai et al.(1999) ⁵⁰	61	27.869	17.147 to 40.829	2.63
Cao et al. (2000) ⁵²	40	72.500	56.112 to 85.399	2.53
Patima et al. (2000) ⁵³	73	73.973	62.376 to 83.546	2.67
Gillison et al.(2000) ⁶⁹	84	11.905	5.859 to 20.805	2.69
Premoli -De-Percoco et al.(2001) ⁵⁴	50	60.000	45.179 to 73.592	2.59
Shima et al.(2000) ⁷⁷	44	20.455	9.804 to 35.305	2.56

Schwartz et al.(2001) ⁵⁵	254	15.748	11.495 to 20.821	2.80
Nagpal et al. (2002) ⁵⁶	110	33.636	24.908 to 43.271	2.73
Kumar et al. (2003) ⁵⁸	42	30.952	17.622 to 47.086	2.54
Sugiyama et al. (2003) ⁶⁰	86	34.884	24.919 to 45.923	2.69
Zhang et al. (2004) ⁶⁴	73	73.973	62.376 to 83.546	2.67
Dahlgren et al. (2004) ⁶²	110	10.909	5.765 to 18.281	2.73
Ibieta et al.(2005) ³⁵	21	66.667	43.032 to 85.412	2.30
Boy et al.(2006) ⁶⁵	59	11.864	4.906 to 22.929	2.63
Badaracco et al.(2006) ⁷⁰	60	13.333	5.936 to 24.592	2.63
Nemes et al.(2006) ⁶⁷	79	41.772	30.767 to 53.413	2.68
Sugiyama et al. (2007) ⁶⁸	27	37.037	19.401 to 57.632	2.40
Smith et al. (2008) ⁷¹	108	24.074	16.368 to 33.251	2.73
Chaudhary et al.(2010) ³⁷	208	34.615	28.171 to 41.505	2.79
Gonzalez-Ramirez I et al.(2013) ³⁴	80	5.000	1.379 to 12.310	2.68
Gan et al.(2014) ³³	200	19.500	14.249 to 25.679	2.79
Koukertsu A et al.(2016) ³⁹	24	54.167	32.821 to 74.447	2.36
Chen et al.(2016) ³⁶	178	3.371	1.247 to 7.192	2.78
Palve et al.(2018) ³⁸	106	33.019	24.190 to 42.824	2.72
De Abreu PM (2018) ⁷⁶	90	3.333	0.693 to 9.434	2.70
Rubab Z et al. (2018) ⁷⁵	100	32.000	23.022 to 42.077	2.72
Total (random effects)	3158	28.877	22.617 to 35.570	100.00

Test for heterogeneity

Q	612.7992
DF	37
Significance level	P < 0.0001
I² (inconsistency)	93.96%
95% CI for I²	92.57 to 95.10

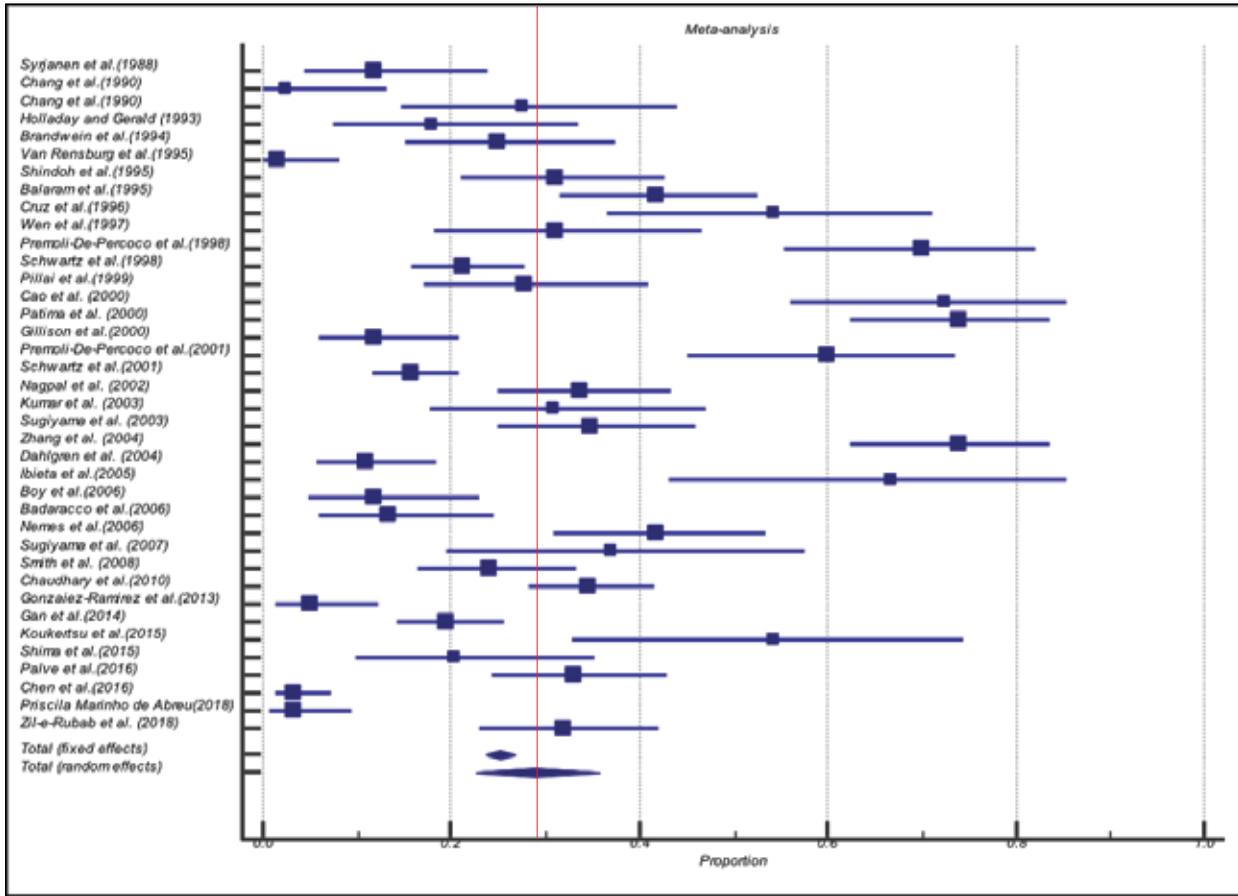


Figure 2: Forest Plot for Prevalence of HPV 16/18 in Oral Cancer.

Meta-analysis for overall prevalence based on 31, 33, 35, 52 and 56 HPV type

Of the 47 research studies, 7 studies included to calculate pooled prevalence of HPV- 31, 33, 35, 52 and 56) in OSCC. The insignificant heterogeneity ($p=0.2294$) was found among the studies [$I^2= 26.11\%$ with 0 to 67.83% confidence interval (CI) at 95%].

Out of 563 patients, 19 patients were positive for HPV-31/33/ 35/ 52/56. Proportion of positive cases for HPV 31/33/ 35/ 52/56 was also analyzed in the present meta-analysis. Based on fixed effect model the mean proportion of positive cases was 3.591% [2.221 to 5.465% confidence interval (CI) at 95%].

Table 3: Prevalence of other high-risk HPV in Oral Cancer based on 7 studies.

Study	Sample size	Proportion (%)	95% CI	Weight (%)
Bouda et al.(2000) ⁵¹	19	5.263	0.133 to 26.028	3.51
Chang et al. (2003) ⁵⁷	103	0.971	0.0246 to 5.291	18.25
Ritchie et al. (2003) ⁵⁹	141	2.837	0.778 to 7. 104	24.91
Smith et al. (2004) ⁶³	106	2.830	0.587 to 8.049	18.77
El -Mofty et al. (2006) ⁶⁶	94	2.128	0.259 to 7.475	16.67
Kong et al.(2009) ⁷²	13	7.692	0.195 to 36.030	2.46
Attner et al.(2010) ⁷³	87	8.046	3.296 to 15.878	15.44
Total (fixed effects)	563	3.591	2.221 to 5.465	100.00

Test for heterogeneity

Q	8.1200
DF	6
Significance level	P = 0.2294
I2 (inconsistency)	26.11%
95% CI for I2	0.00 to 67.83

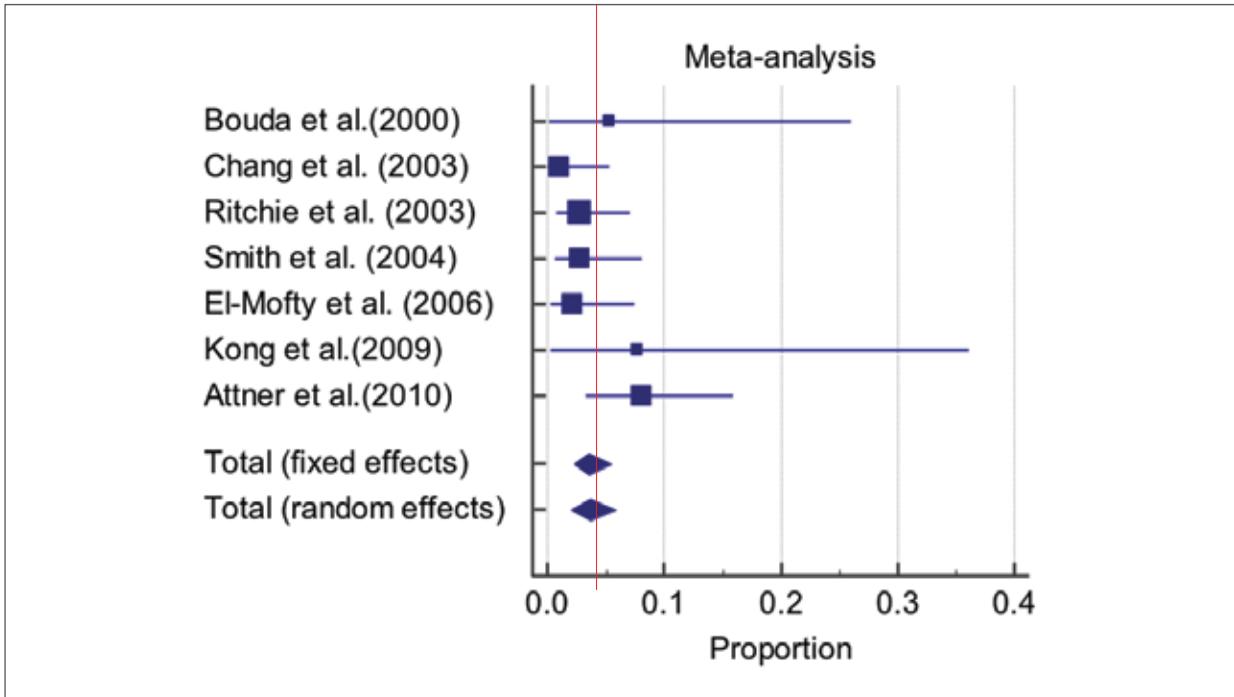


Figure 3: Forest Plot for Prevalence of HPV 31/33/ 35/ 52/56 in Oral Cancer.

DISCUSSION

The present study was designed to analyze the existing highly varied and controversial data on high-risk HPV detected in OSCC in order to calculate the pooled prevalence rate of high-risk HPV and especially HPV 16/18 in oral squamous Cell Carcinoma cases by meta-analysis. The consensus decision supports the moderate prevalence of high-risk HPV type as 30.71% depicting a definite role of high risk HPV in oral oncogenes. Moreover, as highly supported in the literature HPV type 16/18 stand out as the predominant risk factors amongst all. Collectively there seems to be an alarming prevalence of HPV in relation to oral cancer signifying an important role in the etiopathogenetic mechanism, which is in conformity with the already established role of HPV 16/18 in cervical oncogenesis. The massive representing from the western world could be related to the promiscuous society and practice of oral sex.

Well, out of all the high-risk viruses HPV 16/18 is most commonly implicated in the oral carcinogenesis model. However, we have also estimated the prevalence

of other less common high-risk HPV types to prompt further investigation in this subject area. Since most of the work available in the literature has focused on the type 16/18 only. So we analyzed this data separately also for the conclusive comment. However, we could not determine a significant role of other high-risk viruses.

The association of HPV with oral cancer was first proposed in 1983¹ and then supported by several other studies. The HPV prevalence rate in oral cancer was found to be varied from 0% to 100%¹³. The studies including large sample size (> 90 cases) have shown low prevalence 3.37% to 49.51% as compared to the studies based on small sample size 1.52% to 94.73% of high-risk HPV amongst OSCC. The reason of this huge widespread variability maybe due to differences in detection methodology of HPV, differences in population, social habits, and sampling of oral specimen¹⁹⁻²¹. Moreover, many research studies have stated inconsistent results for the prognostic role of HPV^{22, 23}. This is still in debate and needs future investigation.

A study reported that serum analysis, histopatholo-

gy examination and OralCDx® (oral scrapings) diagnostic methods are significant in identifying the HPV DNA. A diagnostic method is considered essential for estimating the prevalence or association of HPV and oral cancer²⁴. According to current study, majority of the research studies used PCR method, which is definitely a standard presently.

Based on our study's statistical results, the overall positive prevalence of high-risk HPV and particularly type 16/18 HPV in OSCC was 30.71% and 28.88% respectively. In addition, 7 out of 47 studies, which determine less than 10% prevalence of all high-risk HPV in OSCC, as, showed in Figure 1. On the contrary, Bouda et al. found high prevalence of 94.74% with 93.69 – 95.71 CI at 95%. While Schwartz et al. studied on the highest number of patients (Table 1). Although, there were only 5 out of 38 studies which determine less than 10% prevalence of HPV16/18 in OSCC, as shown in Figure 2. On the contrary, Patima et al. and Zhang et al. found high prevalence of 73.97% with 62.37 to 83.54% and 62.37 to 83.54% CI at 95% respectively. This large variation in the data requires future rigorous investigation of research derives with good number of cases and uniformity of standardized molecular genetics techniques.

Furthermore, according to earlier published systematic review and meta-analysis our result was almost similar to that stated by Syrjänen et al. 33.7%²⁵ although our result was greater than Miller and White 26.2%²⁶, Ndiaye et al. 24.2%²⁷, Kreimer et al. 23.5%²⁸, and O'Rorke et al. 22.8%²⁹. Whereas our result was, lower than Termine et al. 38.1%³, Miller and Johnstone 46.5%³⁰, Chaitanya et al. 58%²⁴.

CONCLUSION

According to the current meta-analysis, we assumed that high infection rate of type 16/18 HPV as compared to other high-risk HPV, which were found in OSCC cases remains a scientific reality in the risk-factor profile.

LIST OF ABBREVIATIONS

OSCC	Oral Squamous Cell Carcinoma
HPV	Human Papillomaviruses
PCR	Polymerase Chain Reaction
ISH	In-situ Hybridization
CI	Confidence Interval
DNA	Deoxyribonucleic Acid
LCR	Long non-Coding Region
ORF	Open Reading Frame
DF	Degree of Freedom

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CONFLICT OF INTEREST

There was no conflict of interest among the authors.

AUTHORS' CONTRIBUTIONS

TM conceived the idea, guided the manuscript, overall supervised the project, SU wrote introduction of the manuscript and facilitated in online data collection, AW collected online data, did meta-analysis and wrote the manuscript.

REFERENCES

1. Markopoulos AK. Current Aspects on Oral Squamous Cell Carcinoma. *Open Dent J.* 2012; 6: 126-130.
2. Dong Y, Zhao Q, Ma X, Ma G, Liu C, Chen Z, Yu L, Liu X, Zhang Y, Shao S, Xiao J. Establishment of a new OSCC cell line derived from OLK and identification of malignant transformation-related proteins by differential proteomics approach. *Sci Rep.* 2015;5:12668.
3. Williams HK. Molecular pathogenesis of oral squamous carcinoma. *Mol Pathol.* 2000; 53: 165-72.
4. Yete S, D'Souza W, Saranath D. High-risk human papillomavirus in oral cancer: clinical implications. *Oncol.* 2018;94(3):133-41.
5. Siddiqui IA, Farooq MU, Siddiqui RA, Rafi SMT. Role of toluidine blue in early detection of oral cancer. *Pak J Med Sci.* 2006; 22:184-7.
6. Choi S, Myers JN. Molecular pathogenesis of oral squamous cell carcinoma: implications for therapy. *J Dent Res.* 2008; 87: 14-32.
7. Leemans CR, Braakhuis BJ, Brakenhoff RH. The molecular biology of head and neck cancer. *Nat Rev Cancer.* 2011; 11 [1]: 9-22.
8. Ogden GR. Alcohol and oral cancer. *Alcohol.* 2005; 35:169-73.
9. Danaei G, Ding EL, Mozaffarian D, Taylor B, Rehm J, Murray C, Ezzati M. The preventable causes of death in the United States; comparative risk assessment of dietary, lifestyle, and metabolic risk factors. *PLOS Med.* 2009; 6, 1000058.
10. Su CC, Yang HF, Huang SJ, Lian IB. Distinctive features of oral cancer in Changhua County: high incidence, buccal mucosa preponderance, and a close relation to betel quid chewing habit. *J Formos Med Assoc.* 2007; 106: 225-33.
11. Thavarajah R, Rao A, Raman U, Rajasekaran ST, Joshua ERH, Kannan R. Oral lesions of 500 habitual psychoactive substance users in Chennai, India. *Arch Oral Biol.* 2006; 51: 512-9.
12. Akhter M, Ali L, Hassan Z, Khan I. Association of human papilloma virus infection and oral squamous cell carcinoma in Bangladesh. *J Health Popul Nutr.* 2013;31(1):65.
13. Shigeishi H, Sugiyama M. Risk factors for oral human papillomavirus infection in healthy individuals: a systematic review and meta-analysis. *J Clin Med Res.* 2016;8(10):721.
14. Smith EM, Ritchie JM, Summersgill KF, Klussmann

- JP, Lee JH, Wang D, Haugen TH, Turek LP: Age, sexual behavior and human papillomavirus infection in oral cavity and oropharyngeal cancers. *Int J Cancer*. 2004; 108: 766–772.
15. Doorbar J, Quint W, Banks L, Bravo IG, Stoler M, Broker TR, Stanley MA. The biology and life-cycle of human papillomaviruses. *Vaccine*. 2012;30:F55-70.
16. Jung WW, Chun T, Sul D, Hwang KW, Kang HS, Lee DJ, Han IK. Strategies against human papillomavirus infection and cervical cancer. *J Microbiol*. 2004;42(4):255-66.
17. Thorland EC, Myers SL, Gostout BS, Smith DI. Common fragile sites are preferential targets for HPV16 integrations in cervical tumors. *Oncogene*. 2003; 22:1225–1237.
18. Marur S, D'Souza G, Westra WH, Forastiere AA: HPV-associated head and neck cancer: a virus-related cancer epidemic. *Lancet Oncol*. 2010; 11: 781–789.
19. Syrjanen KJ, Pyrhonen S, Syrjanen SM, Lamberg MA. Immunohistochemical demonstration of human papilloma virus (HPV) antigens in oral squamous cell lesions. *Br J Oral Surg*. 1983;21(2):147–53.
20. Castro TP, Bussoloti Filho I. Prevalence of human papillomavirus (HPV) in oral cavity and oropharynx. *Braz J Otorhinolaryngol*. 2006;72(2):272–82.
21. Termine N, Panzarella V, Falaschini S, Russo A, Matranga D, Lo Muzio L, et al. HPV in oral squamous cell carcinoma vs head and neck squamous cell carcinoma biopsies: a meta-analysis (1988-2007). *Ann Oncol*. 2008;19(10):1681–90.
22. Chien CY, Su CY, Fang FM, Huang HY, Chuang HC, Chen CM, et al. Lower prevalence but favorable survival for human Papillomavirus related squamous cell carcinoma of tonsil in Taiwan. *Oral Oncol*. 2008;44(2):174–9.
23. Marques-Silva L, Farias LC, Fraga CA, de Oliveira MV, Cardos CM, Fonseca-Silva T, et al. HPV-16/18 detection does not affect the prognosis of head and neck squamous cell carcinoma in younger and older patients. *Oncol Lett*. 2012;3(4):945–9.
24. Chaitanya NC, Allam NS, Gandhi Babu DB, Waghay S, Badam RK, Lavanya R. Systematic meta-analysis association of human papilloma virus and oral cancer. *J Can Res Ther*. 2016;12:969-74.
25. Syrjanen S, Lodi G, von Bultzingslowen I, Aliko A, Arduino P, Campisi G, Challacombe S, Ficarra G, Flaitz C, Zhou HM, Maeda H: Human papillomaviruses in oral carcinoma and oral potentially malignant disorders: a systematic review. *Oral Dis*. 2011; 17: 58–72.
26. Miller CS, White DK: Human Papillomavirus expression in oral mucosa, premalignant conditions, and squamous cell carcinoma: a retrospective review of the literature. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 1996; 82: 57–68.
27. Ndiaye C, Mena M, Alemany L, Arbyn M, Castellsague X, Laporte L, Bosch FX, de Sanjose S, Trottier H: HPV DNA, E6/E7 mRNA, and p16 INK4a detection in head and neck cancers: a systematic review and meta-analysis. *Lancet Oncol*. 2014; 15: 1319–1331.
28. Kreimer AR, Clifford GM, Boyle P, Franceschi S. Human papillomavirus types in head and neck squamous cell carcinomas worldwide: a systematic review. *Cancer Epidemiol Biomarkers Prev*. 2005; 14: 467–475.
29. O'Rorke MA, Ellison MV, Murray LJ, Moran M, James J, Anderson LA. Human Papillomavirus related head and neck cancer survival: a systematic review and meta-analysis. *Oral Oncol*. 2012; 48: 1191–1201.
30. Miller CS, Johnstone BM: Human Papillomavirus as a risk factor for oral squamous cell carcinoma: a meta-analysis, 1982–1997. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2001; 91: 622–635.
31. Balaram P, Nalinakumari KR, Abraham E, Balan A, Hareendran NK, Bernard HU, et al. Human papillomaviruses in 91 oral cancers from Indian betel quid chewers – High prevalence and multiplicity of infections. *Int J Cancer*. 1995;61:450-4.
32. Agrawal GP, Joshi PS, Agrawal A. Role of HPV-16 in pathogenesis of oral epithelial dysplasia and oral squamous cell carcinoma and correlation of p16INK4a expression in HPV-16 positive cases: An immunohistochemical study. *ISRN Pathol*. 2013;2013:1-7.
33. Gan LL, Zhang H, Guo JH, Fan MW. Prevalence of human papillomavirus infection in oral squamous cell carcinoma: A case-control study in Wuhan, China. *Asian Pac J Cancer Prev*. 2014;15:5861-5.
34. Gonzalez-Ramirez I, Irigoyen-Camacho ME, Ramirez-Amador V, Lizano-Soberon M, Carrillo-Garcia A, Garcia-Carranca A, et al. Association between age and high-risk human papillomavirus in Mexican oral cancer patients. *Oral Dis*. 2013;19: 796-804.
35. Ibieta BR, Lizano M, Fras-Mendivil M, Barrera JL, Carrillo A, Ma Ruz-Godoy L, et al. Human papillomavirus in oral squamous cell carcinoma in a Mexican population. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2005;99:311-5.
36. Chen F, Yan L, Liu F, Huang J, Liu F, Wu J, et al. Oral human Papillomavirus infection, sexual behaviors and risk of oral squamous cell carcinoma in Southeast of China: A case control study. *J Clin Virol*. 2016;85:7-12.
37. Chaudhary AK, Pandya S, Mehrotra R, Bharti AC, Singh M, Singh M. Comparative study between the Hybrid Capture II test and PCR based assay for the detection of human papillomavirus DNA in oral submucous fibrosis and oral squamous cell carcinoma. *Virology*. 2010;7:253.
38. Palve V, Bagwan J, Krishnan NM, Pareek M, Chandola U, Suresh A, Siddappa G, James BL, Kekatpure V, Kuriakose MA, Panda B. Detection of high-risk human papillomavirus in oral cavity squamous cell carcinoma using multiple analytes and their role in patient survival. *J Glob Oncol*. 2018;4:1-33.
39. Kouketsu A, Sato I, Abe S, Oikawa M, Shimizu Y, Takahashi T, et al. Detection of human papillomavirus infection in oral squamous cell carcinoma: A

- cohort study of Japanese patients. *J Oral Pathol Med.* 2016;45:565-72.
40. Syrjanen SM, Syrjanen KJ, Happonen RP. Human papillomavirus (HPV) DNA sequences in oral precancerous lesions and squamous cell carcinoma demonstrated by in situ hybridization. *J Oral Pathol.* 1988; 17: 273–278.
 41. Chang F, Syrjanen S, Nuutinen J et al. Detection of human papillomavirus (HPV) DNA in oral squamous cell carcinomas by in situ hybridization and polymerase chain reaction. *Arch Dermatol Res.* 1990; 282: 493–497.
 42. Holladay EB, Gerald WL. Viral gene detection in oral neoplasms using the polymerase chain reaction. *Am J Clin Pathol.* 1993; 100: 36–40.
 43. Brandwein M, Zeitlin J, Nuovo GJ et al. HPV detection using 'hot start' polymerase chain reaction in patients with oral cancer: a clinic-pathological study of 64 patients. *Mod Pathol.* 1994; 7: 720–727.
 44. Shindoh M, Chiba I, Yasuda M et al. Detection of human papillomavirus DNA sequences in oral squamous cell carcinomas and their relation to p53 and proliferating cell nuclear antigen expression. *Cancer.* 1995; 76: 1513–1521.
 45. Van Rensburg EJ, van Heerden WF, Venter EH, Raubenheimer EJ. Detection of human papillomavirus DNA with in situ hybridisation in oral squamous carcinoma in a rural black population. *S Afr Med J.* 1995; 85: 894–896.
 46. Cruz IB, Snijders PJ, Steenbergen RD et al. Age-dependence of human papillomavirus DNA presence in oral squamous cell carcinomas. *Eur J Cancer B Oral Oncol.* 1996; 32B: 55–62.
 47. Wen S, Tsuji T, Li X et al. Detection and analysis of human papillomavirus 16 and 18 homologous DNA sequences in oral lesions. *Anticancer Res.* 1997; 17:307–311.
 48. Premoli-De-Percoco G, Ramirez JL, Galindo I. Correlation between HPV types associated with oral squamous cell carcinoma and cervicovaginal cytology: an in situ hybridization study. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 1998; 86: 77–81.
 49. Schwartz SM, Daling JR, Doody DR et al. Oral cancer risk in relation to sexual history and evidence of human papillomavirus infection. *J Natl Cancer Inst.* 1998; 90: 1626–1636.
 50. Pillai MR, Phanidhara A, Kesari AL et al. Cellular manifestations of human papillomavirus infection in the oral mucosa. *J Surg Oncol.* 1999; 71: 10–15.
 51. Bouda M, Gorgoulis VG, Kastrinakis NG et al. "High risk" HPV types are frequently detected in potentially malignant and malignant oral lesions, but not in normal oral mucosa. *Mod Pathol.* 2000; 13: 644–653.
 52. Cao J, Zhang ZY, Patima et al. Human papillomavirus infection and p53 alteration in oral squamous cell carcinoma. *Chin J Dent Res.* 2000; 3:44–49.
 53. Patima Cao J, Chen WT, Zhang ZY. Detection of high-risk human papillomavirus DNA in oral squamous cell carcinoma. *Shanghai Kou Qiang Yi Xue.* 2000; 9: 212–215.
 54. Premoli-De-Percoco G, Ramirez JL. High risk human papillomavirus in oral squamous carcinoma: evidence of risk factors in a Venezuelan rural population. Preliminary report. *J Oral Pathol Med.* 2001; 30: 355–361.
 55. Schwartz SR, Yueh B, McDougall JK et al. Human papillomavirus infection and survival in oral squamous cell cancer: a population-based study. *Otolaryngol Head Neck Surg.* 2001; 125: 1–9.
 56. Nagpal JK, Patnaik S, Das BR. Prevalence of high-risk human papilloma virus types and its association with P53 codon 72 polymorphism in tobacco addicted oral squamous cell carcinoma (OSCC) patients of Eastern India. *Int J Cancer.* 2002; 97: 649–653.
 57. Chang JY, Lin MC, Chiang CP. High-risk human papillomaviruses may have an important role in non-oral habits-associated oral squamous cell carcinomas in Taiwan. *Am J Clin Pathol.* 2003;120: 909–916.
 58. Kumar RV, Kadkol SS, Daniel R et al. Human papillomavirus, p53 and cyclin D1 expression in oropharyngeal carcinoma. *Int J Oral Maxillofac Surg.* 2003; 32:539–543.
 59. Ritchie JM, Smith EM, Summersgill KF et al. Human papillomavirus infection as a prognostic factor in carcinomas of the oral cavity and oropharynx. *Int J Cancer.* 2003; 104: 336–344.
 60. Sugiyama M, Bhawal UK, Dohmen T et al. Detection of human papillomavirus-16 and HPV-18 DNA in normal, dysplastic, and malignant oral epithelium. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2003; 95: 594–600.
 61. Correnti M, Rivera H, Cavazza ME. Detection of human papillomaviruses of high oncogenic potential in oral squamous cell carcinoma in a Venezuelan population. *Oral Dis.* 2004; 10: 163–166.
 62. Dahlgren L, Dahlstrand HM, Lindquist D et al. Human papillomavirus is more common in base of tongue than in mobile tongue cancer and is a favorable prognostic factor in base of tongue cancer patients. *Int J Cancer.* 2004; 112:1015–1019.
 63. Smith EM, Ritchie JM, Summersgill KF et al. Age, sexual behavior and human papillomavirus infection in oral cavity and oropharyngeal cancers. *Int J Cancer.* 2004; 108: 766–772.
 64. Zhang ZY, Sdek P, Cao J, Chen WT. Human papillomavirus type 16 and 18 DNA in oral squamous cell carcinoma and normal mucosa. *Int J Oral Maxillofac Surg.* 2004; 33: 71–74.
 65. Boy S, Van Rensburg EJ, Engelbrecht S et al. HPV detection in primary intraoral squamous cell carcinomas—commensal, aetiological agent or contamination? *J Oral Pathol Med.* 2006; 35: 86–90.
 66. El-Mofty SK, Patil S. Human papillomavirus (HPV)-related oropharyngeal non keratinizing squamous cell carcinoma: characterization of a distinct phenotype. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2006; 101:339–345.
 67. Nemes JA, Deli L, Nemes Z, Marton IJ. Expression

of p16 (INK4A), p53, and Rb proteins are independent from the presence of human papillomavirus genes in oral squamous cell carcinoma. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2006; 102: 344–352.

68. Sugiyama M, Bhawal UK, Kawamura M et al. Human papillomavirus-16 in oral squamous cell carcinoma: clinical correlates and 5-year survival. *Br J Oral Maxillofac Surg.* 2007; 45: 116–122.

69. Gillison ML, Koch WM, Capone RB, Spafford M, Westra WH. Evidence for a causal association between human papillomavirus and a subset of head and neck cancers. *J Natl Cancer Inst.* 2000;92:702–20.

70. Badaracco G, Rizzo C, Mafera B, Pichi B, Giannarelli D, Rahimi SS, et al. Molecular analyses and prognostic relevance of HPV in head and neck tumours. *Oncol Rep.* 2007;17:931–9.

71. Smith EM, Wang D, Rubenstein LM, Morris WA, Turek LP, Haugen TH. Association between p53 and human papillomavirus in head and neck cancer survival. *Cancer Epidemiol Biomarkers Prev.* 2008;17:421–7.

72. Kong CS, Narasimhan B, Hongbin C, Kwok S, Erickson JP, Koong A, et al. The relationship between human papillomavirus status and other molecular prognostic markers in head and neck

squamous cell carcinomas. *Int J Radiat Oncol Biol Phys.* 2009;74:553–61.

73. Attner P, Du J, Nasman A, Hammarstedt L, Ramqvist T, Lindholm J, et al. The role of human papillomavirus in the increased incidence of base of tongue cancer. *Int J Cancer.* 2010;126:2879–84.

74. Chiba I, Shindoh M, Yasuda M, Yamazaki Y, Amemiya A, Sato Y, et al. Mutations in the p53 gene and human papillomavirus infection as significant prognostic factors in squamous cell carcinomas of the oral cavity. *Oncogene.* 1996; 12:1663–8.

75. Rubab Z, Baig S, Zaman U, Lucky MH. Human papilloma virus 16/18: Fabricator of trouble in oral squamous cell carcinoma. *Int J Infect Dis.* 2018; 69:115-119.

76. De Abreu PM, C6 ACG, Azevedo PL, do Valle IB, de Oliveira KG, Gouvea SA, Cordeiro-Silva MF, Louro ID, de Podest6 JRV, Lenzi J, Sena A, Mendonça EF, von Zeidler SLV. Frequency of HPV in oral cavity squamous cell carcinoma. *BMC Cancer.* 2018; 18(1):324.

77. Shima K, Kobayashi I, Saito I, Kiyoshima T, Matsuo K, Ozeki S, et al. Incidence of human papillomavirus 16 and 18 infection and p53 mutation in patients with oral squamous cell carcinoma in Japan. *Br J Oral Maxillofac Surg* 2000; 38:445-50.

